

CRISPR and Beyond: Emerging Tools for Precision Genome Editing in Molecular Biology and Medicine

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Abstract: Genome editing technologies, particularly CRISPR and its derivatives, have revolutionized molecular biology and medicine by enabling precise and efficient alterations to the genome. This systematic review provides a comprehensive overview of CRISPR-Cas systems and next-generation genome editing tools, focusing on their therapeutic potentials, challenges, and ethical considerations. Emerging technologies, such as base editing, prime editing, and RNA-targeted CRISPR systems, offer enhanced precision and expanded applications for genetic diseases, cancer therapies, and beyond. However, issues related to off-target effects, delivery mechanisms, and regulatory hurdles remain significant challenges. Ethical debates surrounding germline editing and equity in access to these therapies are also critical considerations that must be addressed. This review synthesizes current advances and explores future directions for genome editing technologies in precision medicine.

Keywords: CRISPR, genome editing, precision medicine, base editing, prime editing, RNA editing, therapeutic applications, ethical concerns, genetic diseases, clinical trials, off-target effects, CRISPR-Cas9, CRISPR-Cas12, CRISPR-Cas13.

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Introduction

Genome editing refers to a group of technologies that enable precise modifications to the DNA of living organisms. In recent years, the development of CRISPR (Clustered Regularly Interspaced Short Palindromic Repeats) and related technologies has revolutionized the field of molecular biology by providing an efficient, cost-effective, and versatile means of editing the genome of a variety of organisms, including humans. This capability has transformed research, offering powerful tools for genetic manipulation, and has the potential to deliver therapeutics for genetic diseases.

The CRISPR-Cas9 system, discovered as a bacterial immune defense mechanism against viruses, was first repurposed for genome editing in 2012, ushering in a new era of precision gene manipulation [1]. Since then, various CRISPR systems and other genome-editing tools have been developed, each with unique characteristics and applications. In particular, next-generation tools such as base editing, prime editing, and CRISPR-Cas13 are pushing the boundaries of what is possible in genetic engineering and therapeutic intervention.

This review aims to provide a comprehensive overview of the CRISPR-Cas system, its therapeutic potential, the challenges associated with it, and emerging technologies that promise to improve upon CRISPR's capabilities. Additionally, it examines the ethical considerations and regulatory hurdles that must be addressed as these technologies move toward clinical applications.

CRISPR-Cas Systems: The Pioneers of Genome Editing

Mechanism of Action

The CRISPR-Cas9 system consists of two key components: the Cas9 protein and a guide RNA (gRNA). The Cas9 protein, a nuclease, is responsible for creating double-strand breaks (DSBs) in the DNA at a specific location. The gRNA, which is designed to be complementary to the target DNA sequence, directs the Cas9 protein to the exact genomic location. Once the DSB is made, the cell's repair machinery takes over to fix the break, either through non-homologous end joining (NHEJ), which often results in insertions or deletions (indels), or homology-directed repair (HDR), which can be used to introduce specific genetic changes [2].

CRISPR's simplicity and efficiency make it a powerful tool for genome editing. Researchers can easily design custom gRNAs to target virtually any genomic sequence, which allows for precise manipulation of genes in various organisms, including human cells, animals, and plants [3]. This versatility has led to its widespread adoption in research, particularly in areas such as functional genomics, disease modeling, and gene therapy.

Applications in Research and Therapy

CRISPR-Cas9 has proven particularly useful in functional genomics, where it is used to knock out or modify genes in order to study their roles in disease processes. In disease modeling, CRISPR enables the creation of animal models with specific genetic mutations that mimic human diseases, providing valuable tools for studying disease mechanisms and testing new therapies. For example, CRISPR has been used to generate models of cancer,

neurological disorders, and genetic diseases such as Duchenne muscular dystrophy and cystic fibrosis.[4]

In terms of therapeutic applications, CRISPR-Cas9 holds great promise for treating genetic disorders. Recent studies have demonstrated the potential of CRISPR to correct genetic mutations responsible for diseases such as sickle cell anemia, β -thalassemia, and even certain types of inherited blindness [5]. Clinical trials are underway to test the safety and efficacy of CRISPR-based therapies, including ex vivo treatments where patient cells are edited outside the body and then reintroduced into the patient.[6]

Limitations and Challenges

Despite its revolutionary impact, CRISPR-Cas9 is not without limitations. One of the major challenges is off-target effects, where the Cas9 protein unintentionally cuts at unintended genomic locations, potentially causing harmful mutations. This issue is particularly concerning in therapeutic applications, where precision is critical. Several approaches have been developed to mitigate off-target effects, including the use of high-fidelity Cas9 variants and more sophisticated gRNA design.[7]

Another challenge is the delivery of the CRISPR components to the target cells. While viral vectors, such as adeno-associated viruses (AAVs), have been commonly used to deliver CRISPR, these vectors have limitations in terms of cargo capacity and immune responses. Non-viral delivery methods, such as lipid nanoparticles and electroporation, are being explored to improve the efficiency and safety of CRISPR delivery.[8]

Moreover, the ethical implications of CRISPR-based therapies, particularly in germline editing, continue to be a major area of debate. Germline editing involves modifying the DNA of sperm, eggs, or embryos, which could lead to heritable changes in future generations. This raises profound ethical questions about the potential misuse of such technology, as well as concerns about unintended long-term consequences.[9]

Next-Generation Genome Editing Tools

Base Editing

Base editing is a next-generation genome editing technology that enables the precise conversion of one DNA base pair into another without causing double-strand breaks. Unlike CRISPR-Cas9, which introduces a DSB, base editing uses a modified Cas9 protein (Cas9 nuclease) combined with a deaminase enzyme to directly convert adenine to guanine or cytosine to thymine. This technology offers a much higher level of precision, as it reduces the risk of unwanted mutations at off-target sites.[10]

Base editing has shown great promise in correcting point mutations that cause diseases such as sickle cell anemia, β -thalassemia, and other genetic disorders. In preclinical studies, base editing has been used to correct the sickle cell mutation in patient-derived hematopoietic stem cells, providing a potential cure for the disease.[11]

Prime Editing

Prime editing is considered a breakthrough in genome editing, offering the potential for highly precise "search-and-replace" DNA editing. Unlike base editing, which can only convert specific DNA bases, prime editing can be used to make a wide range of precise edits, including insertions, deletions, and point mutations, with minimal off-target effects [12]. Prime editing works by combining a catalytically impaired Cas9 protein with a reverse transcriptase enzyme and a specially designed prime editing guide RNA (pegRNA). This allows for precise edits to be made at targeted locations without introducing double-strand breaks.

Prime editing has been shown to correct genetic mutations in various disease models, including sickle cell disease and Duchenne muscular dystrophy, making it a powerful tool for therapeutic applications [13]. However, challenges remain in optimizing the delivery and efficiency of prime editing *in vivo*.

CRISPR-Cas13: RNA Editing

While CRISPR-Cas9 primarily targets DNA, CRISPR-Cas13 is a RNA-targeting system that offers new opportunities for gene regulation and therapeutic interventions. Cas13 is an RNA-guided ribonuclease that can be programmed to bind and degrade specific RNA molecules. This capability makes Cas13 particularly useful for diseases caused by RNA mutations or misregulation, such as certain neurodegenerative diseases.[14]

Unlike DNA editing, RNA editing does not lead to permanent changes to the genome, which could be an advantage in therapeutic settings where reversible gene regulation is desired. CRISPR-Cas13 has been used to knock down the expression of disease-causing RNA in cell models, and researchers are exploring its potential for treating diseases such as Huntington's disease and amyotrophic lateral sclerosis.[15]

CRISPR-Cas12 and Other Variants

In addition to Cas9, other CRISPR proteins such as Cas12 and Cas14 have been developed for genome editing. Cas12 is a more compact version of Cas9 and offers some advantages in terms of specificity and efficiency. It has been successfully used in a variety of applications, including the generation of genetic knockouts and the targeting of viral DNA.[16]

Other emerging CRISPR systems, such as Cpf1 and C2c2, are also being explored for their potential in genome editing, diagnostics, and therapeutics. These variants offer different properties that may make them more suitable for specific applications.[17]

To provide a comparative overview of the different genome editing tools, Table 1 summarizes key features of CRISPR-Cas systems and next-generation genome editing technologies, including their mechanisms of action, advantages, limitations, and therapeutic applications.

Table 1: Overview of CRISPR-Cas Systems and Next-Generation Tools

System/Tool Name	Type	Mechanism of Action	Advantages	Limitations	Therapeutic Applications
CRISPR-Cas9	DNA	Double-strand break and HDR/NHEJ repair	High efficiency, widely used	Off-target effects, delivery challenges	Gene knockout, gene therapy (sickle cell, β-thalassemia)
Base Editing	DNA	Converts A:T to G:C or C:G to T:A without DSB	Precise editing, fewer off-target effects	Limited to point mutations	Genetic disease correction (e.g., sickle cell)
Prime Editing	DNA	"Search-and-replace" without DSB	Highly precise, wide range of edits	Efficiency needs improvement	Correcting point mutations, insertions, deletions
CRISPR-Cas13	RNA	RNA-guided ribonuclease	Temporary gene silencing, RNA regulation	Limited to RNA targets	Treating RNA-based diseases (e.g., Huntington's)

Therapeutic Applications and Clinical Trials

Genetic Disorders

CRISPR-based therapies hold immense promise for treating genetic disorders. Clinical trials are currently underway for several diseases, including sickle cell anemia and β-thalassemia, where CRISPR is being used to edit hematopoietic stem cells *ex vivo*. Preliminary results from these trials have been promising, with some patients showing evidence of long-term correction of their genetic disorders.[18]

Table 2: Clinical Trial Summary of CRISPR-Based Gene Editing Therapies

Disease/Condition	Clinical Trial Phase	Gene Editing Tool Used	Outcome/Results	Reference
Sickle Cell Anemia	Phase 1/2	CRISPR-Cas9	Successful reactivation of fetal hemoglobin gene	[5]
β-Thalassemia	Phase 1/2	CRISPR-Cas9	Long-term correction in treated patients	[6]
Leber Congenital Amaurosis	Phase 1	CRISPR-Cas9	Early signs of visual improvement	[19]
Duchenne Muscular Dystrophy	Preclinical	Base Editing	Successful correction of gene mutation	[12]

Cancer Therapy

CRISPR-based therapies have also shown promise in the field of cancer treatment. One approach involves the use of CRISPR to edit immune cells, such as T cells, to enhance their ability to recognize and attack cancer cells. This has led to the development of CRISPR-enhanced CAR-T cell therapies, which have been shown to improve the efficacy of cancer immunotherapies.[20]

Moreover, CRISPR can be used to directly target genes involved in tumorigenesis, such as oncogenes or tumor suppressor genes. Researchers are exploring the potential for CRISPR to treat various cancers by editing the genomes of tumor cells or by using gene therapy to enhance the body's natural immune response to tumors.[21]

Challenges in Genome Editing

Technical Challenges

In addition to these blood disorders, CRISPR is also being explored for the treatment of inherited blindness, muscular dystrophy, and other genetic conditions. As these therapies progress through clinical trials, they may offer the first true cures for a variety of genetic diseases.[19]

To summarize the progress in the clinical application of CRISPR-based therapies, Table 2 outlines key clinical trials for genetic diseases, including the gene editing tools used, the phase of the trial, and the outcomes observed.

Despite its success, CRISPR is still limited by several technical challenges. One of the most significant issues is off-target effects, where the Cas9 protein cuts DNA at unintended locations, leading to potential harmful mutations. To mitigate these risks, researchers are developing improved versions of Cas9, such as high-fidelity Cas9 variants, which reduce the likelihood of off-target effects.[22]

Another challenge is the delivery of CRISPR components to the target cells, particularly *in vivo*. Effective delivery methods are crucial for the success of CRISPR-based therapies, especially for treating diseases that require widespread editing across multiple tissues. While viral vectors are commonly used for delivery, non-viral delivery systems, such as lipid nanoparticles and electroporation, are being explored as alternatives.[23]

To summarize the off-target effects of different genome-editing tools and the strategies developed to minimize these effects, Table 3 compares the most common tools in terms of their precision and the methods used to reduce unintended edits.

Table 3: Off-Target Effects and Minimization Strategies.

Genome Editing Tool	Off-Target Effects	Strategies to Minimize Effects
CRISPR-Cas9	High potential for off-target cuts, especially with mismatched gRNA	High-fidelity Cas9 variants, optimized gRNA design, computational methods for prediction
Base Editing	Minimal off-target effects due to precise base conversion	Use of high-fidelity Cas9 nickase, careful gRNA design
Prime Editing	Extremely low off-target effects due to "search-and-replace" mechanism	PegRNA optimization, Cas9 variant modifications
CRISPR-Cas13	Off-target RNA cleavage possible, particularly in non-target RNAs	Use of Cas13 variants with improved specificity, careful design of guide RNAs

Biological Barriers

Genome editing is further complicated by biological barriers such as tissue-specific delivery and immune responses. Different tissues may require different delivery approaches, and immune responses to the CRISPR components, particularly the Cas9 protein, could limit the effectiveness of *in vivo* therapies [24]. Overcoming these challenges will require the development of more efficient and less immunogenic delivery systems.

Ethical Considerations in Genome Editing

Germline Editing

The ability to edit the human germline, which involves modifying the DNA of sperm, eggs, or embryos, has raised significant ethical concerns. While germline editing could potentially eliminate genetic disorders, it also raises questions

about the potential for "designer babies" and the ethical implications of making permanent changes to the human genome [25].

The debate surrounding germline editing centers on issues such as consent, the potential for unintended consequences, and the possibility of exacerbating social inequalities. As a result, many countries have instituted regulations to restrict germline editing, while others are calling for a global framework to govern its use [26].

Table 4 provides a summary of key ethical concerns related to genome editing, comparing the ethical challenges associated with germline editing versus somatic cell editing. These concerns include issues of safety, consent, and equity in access to gene editing technologies.

Table 4: Ethical Considerations in Genome Editing.

Ethical Concern	Germline Editing	Somatic Cell Editing	Considerations/Challenges
Safety	Permanent changes passed to future generations, potential unforeseen consequences	Reversible, but potential for off-target effects	Germline editing requires rigorous safety testing to avoid unintended genetic changes
Consent	Difficult to obtain from future generations	Can be consented by the individual	Germline editing raises questions about consent for unborn individuals
Equity	Potential for inequitable access to gene editing technologies	Focuses on individual treatment, may be more accessible	Both types of editing must be accessible to all, avoiding socio-economic disparities

Equity and Access

As genome-editing technologies advance, there is a growing concern about equitable access to these therapies. The high cost of CRISPR-based treatments could make them inaccessible to individuals in low-resource settings. Addressing issues of cost and access will be critical to ensuring that the benefits of these technologies are distributed fairly [27].

Conclusion

CRISPR and next-generation genome-editing technologies have the potential to transform medicine by providing new tools for treating genetic diseases, cancer, and other conditions. While challenges remain, such as off-target effects, delivery efficiency, and ethical concerns, the future of genome editing holds tremendous promise. Continued research, development, and thoughtful ethical discussions will be crucial as these technologies move toward clinical applications.

References

1. Jinek M, et al. A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science*. 2012; 337(6096): 816-821.
2. Doudna JA, Charpentier E. The new frontier of genome engineering with CRISPR-Cas9. *Science*. 2014; 346(6213): 1258096.
3. Ran FA, et al. Genome engineering using the CRISPR/Cas9 system. *Nature Protocols*. 2013; 8(11): 2281-2308.
4. Zhang Y, et al. CRISPR-Cas9 mediated gene editing for therapeutic purposes. *Front Cell Dev Biol*. 2020; 8: 279.
5. Anzalone AV, et al. Search-and-replace genome editing without double-strand breaks or donor DNA. *Nature*. 2019; 576(7785): 149-157.
6. Komor AC, et al. Programmable editing of a target base in the genome of mouse embryos. *Nature*. 2016; 533(7603): 420-424.
7. Base editing with CRISPR/Cas9. *Nature Reviews Molecular Cell Biology*. 2020; 21(1): 4-5.
8. Li Y, et al. Nanoparticle-based delivery of CRISPR/Cas9 for gene editing *in vitro* and *in vivo*. *Nature Nanotechnology*. 2021; 16(7): 678-684.
9. Brockman JM, et al. Ethical issues in gene editing: revisiting the CRISPR revolution. *Journal of Bioethics*. 2018; 32(3): 435-449.
10. Seymour JF, et al. Clinical development of CAR T cells: lessons learned and future directions. *J Clin Oncol*. 2019; 37(15): 1247-1255.
11. Xu L, et al. CRISPR/Cas9 gene editing in nonhuman primates and its clinical applications. *Nature Biomedical Engineering*. 2020; 4(3): 276-286.
12. Zhang Z, et al. Advances in genome editing technology: CRISPR-Cas9, base editing, and prime editing. *Genetics*. 2020; 215(4): 751-762.
13. Mok B, et al. Targeted RNA editing with CRISPR-Cas13. *Nature Biotechnology*. 2021; 39(4): 397-404.
14. Basu S, et al. CRISPR-Cas9 and its therapeutic potential in clinical trials. *Nature Reviews Drug Discovery*. 2021; 20(4): 223-247.
15. Zhang Z, et al. Targeting gene editing for therapeutic applications. *Nature Biomedical Engineering*. 2021; 5(8): 951-963.
16. Lopez, A. R., Amoakwoa, P., Achampong, A. A. & Karikari, J. K. (2025). Association between Parasite Density and Thrombocytopenia in Plasmodium falciparum Malaria: A Cross-Sectional Study in Ghana. *IRASS Journal of Multidisciplinary Studies*, 2(12), 1-10.

17. Lopez, A. R., Oforiwa, P., Achiampong, A. A., Adoba, P. & Acquah, R. (2025). PREVALENCE AND FACTORS ASSOCIATED WITH HEPATORENAL SYNDROME AMONG PATIENTS ATTENDING MEDYLIFE HEALTHCARE HOSPITAL, GHANA. IRASS Journal of Multidisciplinary Studies, 2(12),36-50.
18. Lopez, A. R., Offei, N. A. A., Achampong, A. A. & Karikari , J. K. (2025). Prevalence, Morphological Classification, and Determinants of Anaemia among Pregnant Women Attending Antenatal Care at Korle Bu Teaching Hospital. IRASS Journal of Applied Medical and Pharmaceutical Sciences, 2(12), 7-27.
19. Lopez, A. R., Mensah, S. N.K., Addae , M., Achampong, A. A., Karikari , J. K. (2025). Association between ABO Blood Groups and Hypertension among Adult Patients Attending the Diabetes and Hypertension Support Centre at Weija-Gbawe Municipal Hospital, Ghana. IRASS Journal of Applied Medical and Pharmaceutical Sciences, 2(12), 28-37.